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NOVEL TNF RECEPTOR DEATH DOMAIN LIGAND PROTEINS AND INHIBITORS OF LIGAND BINDING

This application is a continuation-in-part of application Ser. No. 08/602,228, filed February 15, 1996, which was a continuation-in-part of application Ser. No. 08/533,901, filed September 26, 1995, which was a continuation-in-part of application Ser. No. 08/494,440, filed June 19, 1995, which was a continuation-in-part of application Ser. No. 08/327,514, filed October 19, 1994.

BACKGROUND OF THE INVENTION

The present invention relates to the field of anti-inflammatory substances and other substances which act by inhibiting binding to the intracellular domain of a tumor necrosis factor receptor (hereinafter "TNF-R"), such as, for example, the P55 type (or TNF-R1) TNF receptor. More particularly, the present invention is directed to novel ligands which bind to the TNF-R intracellular domain and to inhibition or modulation of signal transduction by this receptor.

Tumor necrosis factor (herein "TNF") is a cytokine which produces a wide range of cellular activities. TNF causes an inflammatory response, which can be beneficial, such as in mounting an immune response to a pathogen, or when overexpressed can lead to other detrimental effects of inflammation.

The cellular effects of TNF are initiated by the binding of TNF to its receptors (TNF-Rs) on the surface of target cells. The isolation of polynucleotides encoding TNF-Rs and variant forms of such receptors has been described in European patent publication Nos. EP 308,378, EP 393,438, EP 433,900, EP 526,905 and EP 568,925; in PCT patent publication Nos. WO91/03553 and WO93/19777; and by Schall *et al.*, Cell *61*:361-370 (1990) (disclosing the P55 type TNF receptor). Processes for purification of TNF-Rs have also been disclosed in U.S. Patent No. 5,296,592.

Native TNF-Rs are characterized by distinct extracellular, transmembrane and intracellular domains. The primary purpose of the extracellular

domain is to present a binding site for TNF on the outside of the cell. When TNF is bound to the binding site, a "signal" is transmitted to the inside of the cell through the transmembrane and intracellular domains, indicating that binding has occurred. Transmission or "transduction" of the signal to the inside of the cell occurs by a change in conformation of the transmembrane and/or intracellular domains of the receptor. This signal is "received" by the binding of proteins and other molecules to the intracellular domain of the receptor, resulting in the effects seen upon TNF stimulation. Two distinct TNF receptors of ~55 kd ("TNF-R1") and ~75 kd ("TNF-R2") have been identified. Numerous studies with anti-TNF receptor antibodies have demonstrated that TNF-R1 is the receptor which signals the majority of the pleiotropic activities of TNF. Recently, the domain required for signaling cytotoxicity and other TNF-mediated responses has been mapped to the ~80 amino acid near the C-terminus of TNF-R1. This domain is therefore termed the "death domain" (hereinafter referred to as "TNF-R death domain" and "TNF-R1-DD") (see, Tartaglia *et al.,* Cell *74*:845-853 (1993)).

While TNF binding by TNF-Rs results in beneficial cellular effects, it is often desirable to prevent or deter TNF binding from causing other detrimental cellular effects. Although substantial effort has been expended investigating inhibition of TNF binding to the extracellular domain of TNF-Rs, examination of binding of proteins and other molecules to the intracellular domain of TNF-Rs has received much less attention.

However, ligands which bind to the TNF-R intracellular domain have yet to be identified. It would be desirable to identify and isolate such ligands to examine their effects upon TNF-R signal transduction and their use as therapeutic agents for treatment of TNF-induced conditions. Furthermore, identification of such ligands would provide a means for screening for inhibitors of TNF-R/intracellular ligand binding, which will also be useful as anti-inflammatory agents.

SUMMARY OF THE INVENTION

Applicants have for the first time identified novel TNF-R1-DD ligand proteins and have isolated polynucleotides encoding such ligands. Applicants have also identified a known protein which may also bind to the death domain of TNF-R.

In one embodiment, the present invention provides a composition comprising an isolated polynucleotide encoding a protein having TNF-R1-DD ligand protein activity. In preferred embodiments, the polynucleotide is selected from the group consisting of:

(a) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:1 from nucleotide 2 to nucleotide 1231;

(b) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:1;

(c) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:2;

(d) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:2;

(e) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:3 from nucleotide 2 to nucleotide 415;

(f) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:3;

(g) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:4;

(h) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:4;

(i) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:9 from nucleotide 2 to nucleotide 931;

(j) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:9;

(k) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:10;

(l) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:10;

(m) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:11 from nucleotide 2 to nucleotide 1822;

(n) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:11;

(o) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:12;

(p) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:12;

(q) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:13 from nucleotide 3 to nucleotide 2846;

(r) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:13, which encodes a protein having TNF-R1-DD ligand protein activity;

(s) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:14;

(t) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:14 and having TNF-R1-DD ligand protein activity;

(u) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:15 from nucleotide 326 to nucleotide 5092;

(v) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:15;

(w) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:16;

(x) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:16;

(y) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:17 from nucleotide 14 to nucleotide 2404;

(z) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:17;

(aa) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:18;

(bb) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:18; and

(cc) a polynucleotide capable of hybridizing under stringent conditions to any one of the polynucleotides specified in (a)-(cc)

In certain preferred embodiments, the polynucleotide is operably linked to an expression control sequence. The invention also provides a host cell, including bacterial, yeast, insect and mammalian cells, transformed with such polynucleotide compositions.

Processes are also provided for producing an TNF-R1-DD ligand protein, which comprises:

(a) growing a culture of the host cell transformed with such polynucleotide compositions in a suitable culture medium; and

(b) purifying the TNF-R1-DD ligand protein from the culture.

The ligand protein produced according to such methods is also provided by the present invention.

Compositions comprising a protein having TNF-R1-DD ligand protein activity are also disclosed. In preferred embodiments the protein comprises an amino acid sequence selected from the group consisting of:

(a) the amino acid sequence of SEQ ID NO:2;

(b) fragments of the amino acid sequence of SEQ ID NO:2;

(c) the amino acid sequence of SEQ ID NO:4;

(d) fragments of the amino acid sequence of SEQ ID NO:4;

(e) the amino acid sequence of SEQ ID NO:6;

(f) fragments of the amino acid sequence of SEQ ID NO:6;

(g) the amino acid sequence of SEQ ID NO:10;

(h) fragments of the amino acid sequence of SEQ ID NO:10;

(i) the amino acid sequence of SEQ ID NO:12;

(j) fragments of the amino acid sequence of SEQ ID NO:12;

(k) the amino acid sequence of SEQ ID NO:14;

(l) fragments of the amino acid sequence of SEQ ID NO:14;

(m) the amino acid sequence of SEQ ID NO:16;

(n) fragments of the amino acid sequence of SEQ ID NO:16;

(o) the amino acid sequence of SEQ ID NO:18; and

(p) fragments of the amino acid sequence of SEQ ID NO:18;

the protein being substantially free from other mammalian proteins. Such compositions may further comprise a pharmaceutically acceptable carrier.

Compositions comprising an antibody which specifically reacts with such TNF-R1-DD ligand protein are also provided by the present invention.

Methods are also provided for identifying an inhibitor of TNF-R death domain binding which comprise:

(a) combining an TNF-R death domain protein with an TNF-R1-DD ligand protein, said combination forming a first binding mixture;

(b) measuring the amount of binding between the TNF-R death domain protein and the TNF-R1-DD ligand protein in the first binding mixture;

(c) combining a compound with the TNF-R death domain protein and an TNF-R1-DD ligand protein to form a second binding mixture;

(d) measuring the amount of binding in the second binding mixture; and

(e) comparing the amount of binding in the first binding mixture with the amount of binding in the second binding mixture;

wherein the compound is capable of inhibiting TNF-R death domain binding when a decrease in the amount of binding of the second binding mixture occurs. In certain preferred embodiments the TNF-R1-DD ligand protein used in such method comprises an amino acid sequence selected from the group consisting of:

(a) the amino acid sequence of SEQ ID NO:2;

(b) fragments of the amino acid sequence of SEQ ID NO:2;

(c) the amino acid sequence of SEQ ID NO:4;

(d) fragments of the amino acid sequence of SEQ ID NO:4;

(e) the amino acid sequence of SEQ ID NO:6;

(f) fragments of the amino acid sequence of SEQ ID NO:6;

(g) the amino acid sequence of SEQ ID NO:8;

(h) fragments of the amino acid sequence of SEQ ID NO:8

(i) the amino acid sequence of SEQ ID NO:10;

(j) fragments of the amino acid sequence of SEQ ID NO:10;

(k) the amino acid sequence of SEQ ID NO:12;

(l) fragments of the amino acid sequence of SEQ ID NO:12;

(m) the amino acid sequence of SEQ ID NO:14;

(n) fragments of the amino acid sequence of SEQ ID NO:14.

(o) the amino acid sequence of SEQ ID NO:16;

(p) fragments of the amino acid sequence of SEQ ID NO:16;

(q) the amino acid sequence of SEQ ID NO:18;

(r) fragments of the amino acid sequence of SEQ ID NO:18.

Compositions comprising inhibitors identified according to such method are also provided. Such compositions may include pharmaceutically acceptable carriers.

Methods are also provided for preventing or ameliorating an inflammatory condition which comprises administering a therapeutically effective amount of a composition comprising a protein having TNF-R1-DD ligand protein activity and a pharmaceutically acceptable carrier.

Other embodiments provide methods of inhibiting TNF-R death domain binding comprising administering a therapeutically effective amount of a composition comprising a protein having TNF-R1-DD ligand protein activity and a pharmaceutically acceptable carrier.

Methods are also provided for preventing or ameliorating an inflammatory condition which comprises administering to a mammalian subject a therapeutically effective amount of a composition comprising a pharmaceutically acceptable carrier and a protein selected from the group consisting of insulin-like growth factor binding protein-5 ("IGFBP-5"), and fragments thereof having TNF-R1-DD ligand protein activity. Such proteins may also be administered for inhibiting TNF-R death domain binding.

Methods of preventing or ameliorating an inflammatory condition or of inhibiting TNF-R death domain binding are provided, which comprise administering to a mammalian subject a therapeutically effective amount of inhibitors of TNF-R death domain binding, are also provided.

Methods of identifying an inhibitor of TNF-R death domain binding are also provided by the present invention which comprise:

(a) transforming a cell with a first polynucleotide encoding an TNF-R death domain protein, a second polynucleotide encoding an TNF-R1-DD ligand protein, and at least one reporter gene, wherein the expression of the reporter gene is regulated by the binding of the TNF-R1-DD ligand protein

encoded by the second polynucleotide to the TNF-R death domain protein encoded by the first polynucleotide;

(b) growing the cell in the presence of and in the absence of a compound; and

(c) comparing the degree of expression of the reporter gene in the presence of and in the absence of the compound;

wherein the compound is capable of inhibiting TNF-R death domain binding when a decrease in the degree of expression of the reporter gene occurs. In preferred embodiments, the cell is a yeast cell and the second polynucleotide is selected from the group consisting of:

(a) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:1 from nucleotide 2 to nucleotide 1231;

(b) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:1, which encodes a protein having TNF-R1-DD ligand protein activity;

(c) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:2;

(d) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:2 and having TNF-R1-DD ligand protein activity;

(e) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:3 from nucleotide 2 to nucleotide 415;

(f) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:3, which encodes a protein having TNF-R1-DD ligand protein activity;

(g) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:4;

(h) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:4 and having TNF-R1-DD ligand protein activity;

(i) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:5 from nucleotide 2 to nucleotide 559;

(j) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:5, which encodes a protein having TNF-R1-DD ligand protein activity;

(k) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:6;

(l) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:6 and having TNF-R1-DD ligand protein activity;

(m) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:7 from nucleotide 57 to nucleotide 875;

(n) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:7, which encodes a protein having TNF-R1-DD ligand protein activity;

(o) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:8;

(p) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:8 and having TNF-R1-DD ligand protein activity;

(q) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:9 from nucleotide 2 to nucleotide 931;

(r) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:9;

(s) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:10;

(t) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:10;

(u) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:11 from nucleotide 2 to nucleotide 1822;

(v) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:11;

(w) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:12;

(x) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:12;

(y) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:13 from nucleotide 3 to nucleotide 2846;

(z) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:13, which encodes a protein having TNF-R1-DD ligand protein activity;

(aa) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:14;

(bb) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:14 and having TNF-R1-DD ligand protein activity;

(cc) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:15 from nucleotide 326 to nucleotide 5092;

(dd) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:15, which encodes a protein having TNF-R1-DD ligand protein activity;

(ee) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:16;

(ff) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:16 and having TNF-R1-DD ligand protein activity;

(gg) a polynucleotide comprising the nucleotide sequence of SEQ ID NO:17 from nucleotide 14 to nucleotide 2404;

(hh) a polynucleotide comprising a fragment of the nucleotide sequence of SEQ ID NO:17, which encodes a protein having TNF-R1-DD ligand protein activity;

(ii) a polynucleotide encoding an TNF-R1-DD ligand protein comprising the amino acid sequence of SEQ ID NO:18;

(jj) a polynucleotide encoding an TNF-R1-DD ligand protein comprising a fragment of the amino acid sequence of SEQ ID NO:18 and having TNF-R1-DD ligand protein activity; and

(kk) a polynucleotide capable of hybridizing under stringent conditions to any one of the polynucleotides specified in (a)-(jj), which encodes a protein having TNF-R1-DD ligand protein activity.

BRIEF DESCRIPTION OF THE FIGURES

Figs. 1 and 2 depict autoradiographs demonstrating the expression of TNF-R1-DD ligand proteins of the present invention.

Fig. 3 depicts an autoradiograph demonstrating the expression of clones 1TU, 15TU and 27TU.

Fig. 4 demonstrates the binding of 1TU and 27TU to TNF-R1-DD. MBP, MBP-1TU or MBP-27TU (3μg) was incubated with glutathione beads containing 3μg of either GST or GST-TNF-R1-DD in 100μl of binding buffer (0.2% Triton, 20 mM Tris pH 7.5, 140 mM NaCl, 0.1 mM EDTA, 10 mM DTT and 5% glycerol). The reaction ws performed at 4°C for 2 hours and centrifuged to remove unbound fraction (Unbound). The beads were then washed with 500μl binding buffer four times and resuspended into SDS-sample buffer (Bound). These samples were analyzed by Western blot using anti-MBP antibody (New England Biolab).

Fig. 5 demonstrates the ability of 15TU and 27TU to activate the JNK pathway. COS cells were contransfected with HA-tagged JNK1 and clones 15tu or 27TU. Cells were left untreated or treated for 15 min with 50 ng/ml TNF, and HA-JNK1 was immunoprecipitated with anti-HA antibody. JNK activity was measured in an *in vitro* kinase assay using GST-c-jun (amino acids 1-79) as substrate, and reactions were electrophoresed on SDS-PAGE.

Fig. 6 is an autoradiograph of an SDS-PAGE gel of conditioned media from COS cells transfected with clone 3TW.

Fig. 7 is an autoradiograph which demonstrates that an antisense oligonucleotide derived from the sequence of clone 3TW inhibits TNF-induced $cPLA_2$ phosphorylation.

DETAILED DESCRIPTION OF THE INVENTION

The present inventors have for the first time identified and isolated novel polynucleotides which encode proteins which bind to the TNF-R death domain. As

used herein "TNF-R" includes all receptors for tumor necrosis factor. The P55 type TNF-R is the preferred receptor for practicing the present invention.

The sequence of a polynucleotide encoding one such protein is set forth in SEQ ID NO:1 from nucleotides 2 to 1231. This polynucleotide has been identified as "clone 2DD" The amino acid sequence of the TNF-R1-DD ligand protein encoded by clone 2DD is set forth in SEQ ID NO:2. It is believed that clone 2DD is a partial cDNA clone of a longer full length coding sequence. However, as demonstrated herein the protein encoded by clone 2DD does bind the death domain of TNF-R (i.e., has "TNF-R1-DD ligand protein activity" as defined herein). Clone 2DD was deposited with the American Type Culture Collection on October 13, 1994 and given the accession number ATCC 69706.

The protein encoded by clone 2DD is 410 amino acids in length. No identical or closely related sequences were found using BLASTN/BLASTX or FASTA searches. Therefore, clone 2DD encodes a novel protein.

The sequence of a polynucleotide encoding one such protein is set forth in SEQ ID NO:3 from nucleotides 2 to 415. This polynucleotide has been identified as "clone 3DD". The amino acid sequence of the TNF-R1-DD ligand protein encoded by clone 3DD is set forth in SEQ ID NO:4. It is believed that clone 3DD is a partial cDNA clone of a longer full length coding sequence. However, as demonstrated herein the protein encoded by clone 3DD does bind the death domain of TNF-R (i.e., has "TNF-R1-DD ligand protein activity" as defined herein). Clone 3DD was deposited with the American Type Culture Collection on October 13, 1994 and given the accession number ATCC 69705.

The protein encoded by clone 3DD is 138 amino acids. No identical or closely related sequences were found using BLASTN/BLASTX or FASTA searches. Therefore, clone 3DD encodes a novel protein.

A full-length clone corresponding to clone 3DD was also isolated and identified as "clone 3TW". The nucleotide sequence of clone 3TW is reported as SEQ ID NO:13. Nucleotides 3 to 2846 of SEQ ID NO:13 encode a TNF-R1-DD ligand protein, the amino acid sequence of which is reported as SEQ ID NO:14. Amino acids 811 to 948 of SEQ ID NO:14 correspond to amino acids 1 to 138 of SEQ ID NO:4

(clone 3DD). Clone 3TW was deposited with the American Type Culture Collection on September 26, 1995 and given the accession number ATCC 69904.

The sequence of a polynucleotide encoding another such protein is set forth in SEQ ID NO:5 from nucleotides 2 to 559. This polynucleotide has been identified as "clone 20DD." The amino acid sequence of the TNF-R1-DD ligand protein encoded by clone 20DD is set forth in SEQ ID NO:6. It is believed that clone 20DD is a partial cDNA clone of a longer full length coding sequence. However, as demonstrated herein the protein encoded by clone 20DD does bind the death domain of TNF-R (*i.e.*, has "TNF-R1-DD ligand protein activity" as defined herein). Clone 20DD was deposited with the American Type Culture Collection on October 13, 1994 and given the accession number ATCC 69704.

The protein encoded by clone 20DD is identical to amino acids 87 to 272 of insulin-like growth factor binding protein-5 ("IGFBP-5"), a sequence for which was disclosed in J. Biol. Chem. *266*:10646-10653 (1991) by Shimasaki *et al.*, which is incorporated herein by reference. The polynucleotide and amino acid sequences of IGFBP-5 are set forth in SEQ ID NO:7 and SEQ ID NO:8, respectively. Based upon the sequence identity between clone 20DD and IGFBP-5, IGFBP-5 and certain fragments thereof will exhibit TNF-R1-DD ligand binding activity (as defined herein).

The sequence of a polynucleotide encoding another such protein is set forth in SEQ ID NO:9 from nucleotides 2 to 931. This polynucleotide has been identified as "clone 1TU" The amino acid sequence of the TNF-R1-DD ligand protein encoded by clone 1TU is set forth in SEQ ID NO:10. It is believed that clone 1TU is a partial cDNA clone of a longer full length coding sequence. However, as demonstrated herein the protein encoded by clone 1TU does bind the death domain of TNF-R (i.e., has "TNF-R1-DD ligand protein activity" as defined herein). Clone 1TU was deposited with the American Type Culture Collection on June 7, 1995 and given the accession number ATCC 69848.

The protein encoded by clone 1TU is 310 amino acids in length. No identical or closely related sequences were found using BLASTN/BLASTX or FASTA searches. Therefore, clone 1TU encodes a novel protein.

The sequence of a polynucleotide encoding another such protein is set forth in SEQ ID NO:11 from nucleotides 2 to 1822. This polynucleotide has been

identified as "clone 27TU" The amino acid sequence of the TNF-R1-DD ligand protein encoded by clone 27TU is set forth in SEQ ID NO:12. It is believed that clone 27TU is a partial cDNA clone of a longer full length coding sequence. However, as demonstrated herein the protein encoded by clone 27TU does bind the death domain of TNF-R (i.e., has "TNF-R1-DD ligand protein activity" as defined herein). Clone 27TU was deposited with the American Type Culture Collection on June 7, 1995 and given the accession number ATCC 69846.

The protein encoded by clone 27TU is 607 amino acids in length. No identical or closely related sequences were found using BLASTN/BLASTX or FASTA searches. Therefore, clone 27TU encodes a novel protein. 27TU may be a longer version of clone 2DD. 2DD encodes the same amino acid sequence (SEQ ID NO:2) as amino acids 198-607 encoded by 27TU (SEQ ID NO:12). The nucleotide sequences of 2DD and 27TU are also identical within this region of identity.

An additional "clone 15TU" was isolated which encoded a portion of the 27TU sequence (approximately amino acids 289-607 of SEQ ID NO:12). Clone 15TU was deposited with the American Type Culture Collection on June 7, 1995 and given the accession number ATCC 69847. 15TU comprises the same nucleotide sequence as 27TU over this region of amino acids.

A full-length clone corresponding to clone 27TU was also isolated and identified as "clone 57TU4A". The nucleotide sequence of clone 57TU4A is reported as SEQ ID NO:15. Nucleotides 336 to 5092 of SEQ ID NO:15 encode a TNF-R1-DD ligand protein, the amino acid sequence of which is reported as SEQ ID NO:146 Amino acids 982 to 1588 of SEQ ID NO:16 correspond to amino acids 1 to 607 of SEQ ID NO:12 (clone 27TU). Clone 57TU4A was deposited with the American Type Culture Collection on February 13, 1996 and given the accession number ATCC 69988.

A full-length clone corresponding to clone 1TU was also isolated and identified as "clone 33-1B". The nucleotide sequence of clone 33-1B is reported as SEQ ID NO:17. Nucleotides 14 to 2404 of SEQ ID NO:17 encode a TNF-R1-DD ligand protein, the amino acid sequence of which is reported as SEQ ID NO:18. Amino acids 488 to 797 of SEQ ID NO:18 correspond to amino acids 1 to 310 of SEQ ID NO:10 (clone 1TU). Clone 33-1B was deposited with the American Type Culture Collection on August 13, 1996 and given the accession number ATCC _____.

Polynucleotides hybridizing to the polynucleotides of the present invention under stringent conditions and highly stringent conditions are also part of the present invention. As used herein, "highly stringent conditions" include, for example, 0.2xSSC at 65°C; and "stringent conditions" include, for example, 4xSSC at 65°C or 50% formamide and 4xSSC at 42°C.

For the purposes of the present application, "TNF-R1-DD ligand protein" includes proteins which exhibit TNF-R1-DD ligand protein activity. For the purposes of the present application, a protein is defined as having "TNF-R1-DD ligand protein activity" when it binds to a protein derived from the TNF-R death domain. Activity can be measured by using any assay which will detect binding to an TNF-R death domain protein. Examples of such assays include without limitation the interaction trap assays and assays in which TNF-R death domain protein which is affixed to a surface in a manner conducive to observing binding, including without limitation those described in Examples 1 and 3. As used herein an "TNF-R death domain protein" includes the entire death domain or fragments thereof.

Fragments of the TNF-R1-DD ligand protein which are capable of interacting with the TNF-R death domain or which are capable of inhibiting TNF-R death domain binding (i.e., exhibit TNF-R1-DD ligand protein activity) are also encompassed by the present invention. Fragments of the TNF-R1-DD ligand protein may be in linear form or they may be cyclized using known methods, for example, as described in H.U. Saragovi, *et al.*, Bio/Technology 10, 773-778 (1992) and in R.S. McDowell, *et al.*, J. Amer. Chem. Soc. 114, 9245-9253 (1992), both of which are incorporated herein by reference. Such fragments may be fused to carrier molecules such as immunoglobulins for many purposes, including increasing the valency of TNF-R1-DD ligand protein binding sites. For example, fragments of the TNF-R1-DD ligand protein may be fused through "linker" sequences to the Fc portion of an immunoglobulin. For a bivalent form of the TNF-R1-DD ligand protein, such a fusion could be to the Fc portion of an IgG molecule. Other immunoglobulin isotypes may also be used to generate such fusions. For example, an TNF-R1-DD ligand protein - IgM fusion would generate a decavalent form of the TNF-R1-DD ligand protein of the invention.

The isolated polynucleotide of the invention may be operably linked to an expression control sequence such as the pMT2 or pED expression vectors disclosed in Kaufman *et al.*, Nucleic Acids Res. 19, 4485-4490 (1991), in order to produce the TNF-R1-DD ligand protein recombinantly. Many suitable expression control sequences are known in the art. General methods of expressing recombinant proteins are also known and are exemplified in R. Kaufman, Methods in Enzymology 185, 537-566 (1990). As defined herein "operably linked" means that the isolated polynucleotide of the invention and the expression control sequence are situated within a vector or cell in such a way that the TNF-R1-DD ligand protein is expressed by a host cell which has been transformed (transfected) with the ligated polynucleotide/expression control sequence.

A number of types of cells may act as suitable host cells for expression of the TNF-R1-DD ligand protein. Host cells include, for example, monkey COS cells, Chinese Hamster Ovary (CHO) cells, human kidney 293 cells, human epidermal A431 cells, human Colo205 cells, 3T3 cells, CV-1 cells, other transformed primate cell lines, normal diploid cells, cell strains derived from in vitro culture of primary tissue, primary explants, HeLa cells, mouse L cells, BHK, HL-60, U937, HaK or Jurkat cells.

The TNF-R1-DD ligand protein may also be produced by operably linking the isolated polynucleotide of the invention to suitable control sequences in one or more insect expression vectors, and employing an insect expression system. Materials and methods for baculovirus/insect cell expression systems are commercially available in kit form from, *e.g.*, Invitrogen, San Diego, California, U.S.A. (the MaxBac® kit), and such methods are well known in the art, as described in Summers and Smith, Texas Agricultural Experiment Station Bulletin No. 1555 (1987), incorporated herein by reference.

Alternatively, it may be possible to produce the TNF-R1-DD ligand protein in lower eukaryotes such as yeast or in prokaryotes such as bacteria. Potentially suitable yeast strains include *Saccharomyces cerevisiae, Schizosaccharomyces pombe, Kluyveromyces* strains, *Candida,* or any yeast strain capable of expressing heterologous proteins. Potentially suitable bacterial strains include *Escherichia coli, Bacillus subtilis, Salmonella typhimurium,* or any bacterial strain capable of expressing heterologous proteins. If the TNF-R1-DD ligand protein is made in yeast or bacteria, it may be

necessary to modify the protein produced therein, for example by phosphorylation or glycosylation of the appropriate sites, in order to obtain the functional TNF-R1-DD ligand protein. Such covalent attachments may be accomplished using known chemical or enzymatic methods.

The TNF-R1-DD ligand protein of the invention may also be expressed as a product of transgenic animals, e.g., as a component of the milk of transgenic cows, goats, pigs, or sheep which are characterized by somatic or germ cells containing a nucleotide sequence encoding the TNF-R1-DD ligand protein.

The TNF-R1-DD ligand protein of the invention may be prepared by culturing transformed host cells under culture conditions suitable to express the recombinant protein. The resulting expressed protein may then be purified from such culture (i.e., from culture medium or cell extracts) using known purification processes, such as gel filtration and ion exchange chromatography. The purification of the TNF-R1-DD ligand protein may also include an affinity column containing the TNF-R death domain or other TNF-R death domain protein; one or more column steps over such affinity resins as concanavalin A-agarose, heparin-toyopearl® or Cibacrom blue 3GA Sepharose®; one or more steps involving hydrophobic interaction chromatography using such resins as phenyl ether, butyl ether, or propyl ether; or immunoaffinity chromatography.

Alternatively, the TNF-R1-DD ligand protein of the invention may also be expressed in a form which will facilitate purification. For example, it may be expressed as a fusion protein, such as those of maltose binding protein (MBP) or glutathione-S-transferase (GST). Kits for expression and purification of such fusion proteins are commercially available from New England BioLab (Beverly, MA) and Pharmacia (Piscataway, NJ), respectively. The TNF-R ligand protein can also be tagged with an epitope and subsequently purified by using a specific antibody directed to such epitope. One such epitope ("Flag") is commercially available from Kodak (New Haven, CT).

Finally, one or more reverse-phase high performance liquid chromatography (RP-HPLC) steps employing hydrophobic RP-HPLC media, e.g., silica gel having pendant methyl or other aliphatic groups, can be employed to further purify the TNF-R1-DD ligand protein. Some or all of the foregoing purification steps, in

various combinations, can also be employed to provide a substantially homogeneous isolated recombinant protein. The TNF-R1-DD ligand protein thus purified is substantially free of other mammalian proteins and is defined in accordance with the present invention as an "isolated TNF-R1-DD ligand protein."

TNF-R1-DD ligand proteins may also be produced by known conventional chemical synthesis. Methods for constructing the proteins of the present invention by synthetic means are known to those skilled in the art. The synthetically-constructed protein sequences, by virtue of sharing primary, secondary or tertiary structural and/or conformational characteristics with TNF-R1-DD ligand proteins may possess biological properties in common therewith, including TNF-R1-DD ligand protein activity. Thus, they may be employed as biologically active or immunological substitutes for natural, purified TNF-R1-DD ligand proteins in screening of therapeutic compounds and in immunological processes for the development of antibodies.

The TNF-R1-DD ligand proteins provided herein also include proteins characterized by amino acid sequences similar to those of purified TNF-R1-DD ligand proteins but into which modification are naturally provided or deliberately engineered. For example, modifications in the peptide or DNA sequences can be made by those skilled in the art using known techniques. Modifications of interest in the TNF-R1-DD ligand protein sequences may include the replacement, insertion or deletion of a selected amino acid residue in the coding sequence. For example, one or more of the cysteine residues may be deleted or replaced with another amino acid to alter the conformation of the molecule. Mutagenic techniques for such replacement, insertion or deletion are well known to those skilled in the art (see, e.g., U.S. Patent No. 4,518,584).

Other fragments and derivatives of the sequences of TNF-R1-DD ligand proteins which would be expected to retain TNF-R1-DD ligand protein activity in whole or in part and may thus be useful for screening or other immunological methodologies may also be easily made by those skilled in the art given the disclosures herein. Such modifications are believed to be encompassed by the present invention.

TNF-R1-DD ligand protein of the invention may also be used to screen for agents which are capable of inhibiting or blocking binding of an TNF-R1-DD ligand protein to the death domain of TNF-R, and thus may act as inhibitors of TNF-R death

domain binding and/or TNF activity. Binding assays using a desired binding protein, immobilized or not, are well known in the art and may be used for this purpose using the TNF-R1-DD ligand protein of the invention. Examples 1 and 3 describe examples of such assays. Appropriate screening assays may be cell-based or cell-free. Alternatively, purified protein based screening assays may be used to identify such agents. For example, TNF-R1-DD ligand protein may be immobilized in purified form on a carrier and binding to purified TNF-R death domain may be measured in the presence and in the absence of potential inhibiting agents. A suitable binding assay may alternatively employ purified TNF-R death domain immobilized on a carrier, with a soluble form of a TNF-R1-DD ligand protein of the invention. Any TNF-R1-DD ligand protein may be used in the screening assays described above.

In such a screening assay, a first binding mixture is formed by combining TNF-R death domain protein and TNF-R1-DD ligand protein, and the amount of binding in the first binding mixture (B_o) is measured. A second binding mixture is also formed by combining TNF-R death domain protein, TNF-R1-DD ligand protein, and the compound or agent to be screened, and the amount of binding in the second binding mixture (B) is measured. The amounts of binding in the first and second binding mixtures are compared, for example, by performing a B/B_o calculation. A compound or agent is considered to be capable of inhibiting TNF-R death domain binding if a decrease in binding in the second binding mixture as compared to the first binding mixture is observed. The formulation and optimization of binding mixtures is within the level of skill in the art. Such binding mixtures may also contain buffers and salts necessary to enhance or to optimize binding, and additional control assays may be included in the screening assay of the invention.

Alternatively, appropriate screening assays may be cell based. For example, the binding or interaction between an TNF-R ligand protein and the TNF-R death domain can be measured in yeast as described below in Examples 1 and 3.

Compounds found to reduce, preferably by at least about 10%, more preferably greater than about 50% or more, the binding activity of TNF-R1-DD ligand protein to TNF-R death domain may thus be identified and then secondarily screened in other binding assays, including *in vivo* assays. By these means compounds having

inhibitory activity for TNF-R death domain binding which may be suitable as anti-inflammatory agents may be identified.

Isolated TNF-R1-DD ligand protein may be useful in treating, preventing or ameliorating inflammatory conditions and other conditions, such as cachexia, autoimmune disease, graft versus host reaction, osteoporosis, colitis, myelogenous leukemia, diabetes, wasting, and atherosclerosis. Isolated TNF-R1-DD ligand protein may be used itself as an inhibitor of TNF-R death domain binding or to design inhibitors of TNF-R death domain binding. Inhibitors of binding of TNF-R1-DD ligand protein to the TNF-R death domain ("TNF-R intracellular binding inhibitors") are also useful for treating such conditions.

The present invention encompasses both pharmaceutical compositions and therapeutic methods of treatment or use which employ isolated TNF-R1-DD ligand protein and/or binding inhibitors of TNF-R intracellular binding.

Isolated TNF-R1-DD ligand protein or binding inhibitors (from whatever source derived, including without limitation from recombinant and non-recombinant cell lines) may be used in a pharmaceutical composition when combined with a pharmaceutically acceptable carrier. Such a composition may also contain (in addition to TNF-R1-DD ligand protein or binding inhibitor and a carrier) diluents, fillers, salts, buffers, stabilizers, solubilizers, and other materials well known in the art. The term "pharmaceutically acceptable" means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredient(s). The characteristics of the carrier will depend on the route of administration. The pharmaceutical composition of the invention may also contain cytokines, lymphokines, or other hematopoietic factors such as M-CSF, GM-CSF, TNF, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, G-CSF, Meg-CSF, stem cell factor, and erythropoietin. The pharmaceutical composition may further contain other anti-inflammatory agents. Such additional factors and/or agents may be included in the pharmaceutical composition to produce a synergistic effect with isolated TNF-R1-DD ligand protein or binding inhibitor, or to minimize side effects caused by the isolated TNF-R1-DD ligand protein or binding inhibitor. Conversely, isolated TNF-R1-DD ligand protein or binding inhibitor may be included in formulations of the particular cytokine, lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent

to minimize side effects of the cytokine, lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent.

The pharmaceutical composition of the invention may be in the form of a liposome in which isolated TNF-R1-DD ligand protein or binding inhibitor is combined, in addition to other pharmaceutically acceptable carriers, with amphipathic agents such as lipids which exist in aggregated form as micelles, insoluble monolayers, liquid crystals, or lamellar layers in aqueous solution. Suitable lipids for liposomal formulation include, without limitation, monoglycerides, diglycerides, sulfatides, lysolecithin, phospholipids, saponin, bile acids, and the like. Preparation of such liposomal formulations is within the level of skill in the art, as disclosed, for example, in U.S. Patent No. 4,235,871; U.S. Patent No. 4,501,728; U.S. Patent No. 4,837,028; and U.S. Patent No. 4,737,323, all of which are incorporated herein by reference.

As used herein, the term "therapeutically effective amount" means the total amount of each active component of the pharmaceutical composition or method that is sufficient to show a meaningful patient benefit, i.e., treatment, healing, prevention or amelioration of an inflammatory response or condition, or an increase in rate of treatment, healing, prevention or amelioration of such conditions. When applied to an individual active ingredient, administered alone, the term refers to that ingredient alone. When applied to a combination, the term refers to combined amounts of the active ingredients that result in the therapeutic effect, whether administered in combination, serially or simultaneously.

In practicing the method of treatment or use of the present invention, a therapeutically effective amount of isolated TNF-R1-DD ligand protein or binding inhibitor is administered to a mammal having a condition to be treated. Isolated TNF-R1-DD ligand protein or binding inhibitor may be administered in accordance with the method of the invention either alone or in combination with other therapies such as treatments employing cytokines, lymphokines or other hematopoietic factors. When co-administered with one or more cytokines, lymphokines or other hematopoietic factors, isolated TNF-R1-DD ligand protein or binding inhibitor may be administered either simultaneously with the cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors, or sequentially. If administered sequentially, the attending physician will decide on the appropriate sequence of administering

isolated TNF-R1-DD ligand protein or binding inhibitor in combination with cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors.

Administration of isolated TNF-R1-DD ligand protein or binding inhibitor used in the pharmaceutical composition or to practice the method of the present invention can be carried out in a variety of conventional ways, such as oral ingestion, inhalation, or cutaneous, subcutaneous, or intravenous injection. Intravenous administration to the patient is preferred.

When a therapeutically effective amount of isolated TNF-R1-DD ligand protein or binding inhibitor is administered orally, isolated TNF-R1-DD ligand protein or binding inhibitor will be in the form of a tablet, capsule, powder, solution or elixir. When administered in tablet form, the pharmaceutical composition of the invention may additionally contain a solid carrier such as a gelatin or an adjuvant. The tablet, capsule, and powder contain from about 5 to 95% isolated TNF-R1-DD ligand protein or binding inhibitor, and preferably from about 25 to 90% isolated TNF-R1-DD ligand protein or binding inhibitor. When administered in liquid form, a liquid carrier such as water, petroleum, oils of animal or plant origin such as peanut oil, mineral oil, soybean oil, or sesame oil, or synthetic oils may be added. The liquid form of the pharmaceutical composition may further contain physiological saline solution, dextrose or other saccharide solution, or glycols such as ethylene glycol, propylene glycol or polyethylene glycol. When administered in liquid form, the pharmaceutical composition contains from about 0.5 to 90% by weight of isolated TNF-R1-DD ligand protein or binding inhibitor, and preferably from about 1 to 50% isolated TNF-R1-DD ligand protein or binding inhibitor.

When a therapeutically effective amount of isolated TNF-R1-DD ligand protein or binding inhibitor is administered by intravenous, cutaneous or subcutaneous injection, isolated TNF-R1-DD ligand protein or binding inhibitor will be in the form of a pyrogen-free, parenterally acceptable aqueous solution. The preparation of such parenterally acceptable protein solutions, having due regard to pH, isotonicity, stability, and the like, is within the skill in the art. A preferred pharmaceutical composition for intravenous, cutaneous, or subcutaneous injection should contain, in addition to isolated TNF-R1-DD ligand protein or binding inhibitor, an isotonic vehicle such as Sodium

Chloride Injection, Ringer's Injection, Dextrose Injection, Dextrose and Sodium Chloride Injection, Lactated Ringer's Injection, or other vehicle as known in the art. The pharmaceutical composition of the present invention may also contain stabilizers, preservatives, buffers, antioxidants, or other additives known to those of skill in the art.

The amount of isolated TNF-R1-DD ligand protein or binding inhibitor in the pharmaceutical composition of the present invention will depend upon the nature and severity of the condition being treated, and on the nature of prior treatments which the patient has undergone. Ultimately, the attending physician will decide the amount of isolated TNF-R1-DD ligand protein or binding inhibitor with which to treat each individual patient. Initially, the attending physician will administer low doses of isolated TNF-R1-DD ligand protein or binding inhibitor and observe the patient's response. Larger doses of isolated TNF-R1-DD ligand protein or binding inhibitor may be administered until the optimal therapeutic effect is obtained for the patient, and at that point the dosage is not increased further. It is contemplated that the various pharmaceutical compositions used to practice the method of the present invention should contain about 0.1 μg to about 100 mg of isolated TNF-R1-DD ligand protein or binding inhibitor per kg body weight.

The duration of intravenous therapy using the pharmaceutical composition of the present invention will vary, depending on the severity of the disease being treated and the condition and potential idiosyncratic response of each individual patient. It is contemplated that the duration of each application of the isolated TNF-R1-DD ligand protein or binding inhibitor will be in the range of 12 to 24 hours of continuous intravenous administration. Ultimately the attending physician will decide on the appropriate duration of intravenous therapy using the pharmaceutical composition of the present invention.

Isolated TNF-R1-DD ligand protein of the invention may also be used to immunize animals to obtain polyclonal and monoclonal antibodies which specifically react with the TNF-R1-DD ligand protein and which may inhibit TNF-R death domain binding. Such antibodies may be obtained using either the entire TNF-R1-DD ligand protein or fragments of TNF-R1-DD ligand protein as an immunogen. The peptide immunogens additionally may contain a cysteine residue at the carboxyl terminus, and are conjugated to a hapten such as keyhole limpet hemocyanin (KLH). Methods for

synthesizing such peptides are known in the art, for example, as in R.P. Merrifield, J. Amer.Chem.Soc. 85, 2149-2154 (1963); J.L. Krstenansky, *et al.*, FEBS Lett. 211, 10 (1987).

Monoclonal antibodies binding to TNF-R1-DD ligand protein or to complex carbohydrate moieties characteristic of the TNF-R1-DD ligand glycoprotein may be useful diagnostic agents for the immunodetection of TNF-R ligand protein.

Neutralizing monoclonal antibodies binding to TNF-R1-DD ligand protein or to complex carbohydrates characteristic of TNF-R1-DD ligand glycoprotein may also be useful therapeutics for both inflammatory conditions and also in the treatment of some forms of cancer where abnormal expression of TNF-R1-DD ligand protein is involved. These neutralizing monoclonal antibodies are capable of blocking the signaling function of the TNF-R1-DD ligand protein. By blocking the binding of TNF-R1-DD ligand protein, certain biological responses to TNF are either abolished or markedly reduced. In the case of cancerous cells or leukemic cells, neutralizing monoclonal antibodies against TNF-R1-DD ligand protein may be useful in detecting and preventing the metastatic spread of the cancerous cells, which may be mediated by the TNF-R1-DD ligand protein.

Due to the similarity of their sequences to the insulin growth factor binding protein ("IGFBP-5") and fragments thereof which bind to the TNF-R death domain are proteins having TNF-R1-DD ligand protein activity as defined herein. As a result, they are also useful in pharmaceutical compositions, for treating inflammatory conditions and for inhibiting TNF-R death domain binding as described above for TNF-R1-DD ligand proteins generally.

EXAMPLE 1
CLONING OF TNF-R DEATH DOMAIN LIGAND PROTEIN ENCODING POLYNUCLEOTIDE

A yeast genetic selection method, the "interaction trap" [Gyuris et al, Cell 75:791-803, 1993, which is incorporated herein by reference], was used to screen WI38 cell cDNA libraries (preparation, see below) for proteins that interact with the death domain of the P55 type 1 TNF receptor (TNF-R1-DD). A polynucleotide encoding amino acids 326 to 413 of the P55 type TNF receptor, TNF-R1-DD, was

obtained via the polymerase chain reaction (PCR) using a grafting method. This TNF-R1-DD DNA was then cloned into pEG202 by BamHI and SalI sites, generating the bait plasmid, pEG202-TNF-R1-DD. This plasmid contains the HIS3 selectable marker, and expression of the bait, the LexA-TNF-R1-DD fusion protein, is from the strong constitutive ADH1 promoter. To create the reporter strain carrying the bait protein, yeast strain EGY48, containing the reporter sequence LexAop-Leu2 in place of the chromosomal LEU2, was transformed with pEG202-TNF-R1-DD and pSH18-34 (Ura+), which carries another reporter sequence, LexAop-lacZ. For screening cDNAs encoding proteins that interact with TNF-R1-DD, the expression vector pJG4-5 (TRP1), containing the WI38 cell cDNA library (see below for the cDNA library construction), was transformed into the above strain (EGY48/pEG202-TNF-R1-DD/pSH18-34) according to the method described by Gietz *et al.*, Nucleic Acids Res., *20:*1425 (1992).

cDNA Library Construction:

WI38 cell cDNA library: Double stranded cDNA was prepared from 3ug of WI38 mRNA using reagents provided by the Superscript Choice System (Gibco/BRL, Gaithersberg, MD) with the following substitutions: the first strand synthesis was primed using an oligo dT/XhoI primer/linker, and the dNTP mix was substituted with a mix containing methyl dCTP (Stratagene, LaJolla, CA). The cDNA was modified at both ends by addition of an EcoRI/NotI/SalI adapter linker and subsequently digested with XhoI. This produced cDNA molecules possessing an EcoRI/NotI/SalI overhang at the 5' end of the gene and an XhoI overhang at the 3' end. These fragments were then ligated into the yeast expression/fusion vector pJG4-5 (Gyuris et al., Cell, 75, 791-803, 1993), which contains at its amino terminus, the influenza virus HA1 epitope tag, the B42 acidic transcription activation domain, and the SV40 nuclear localization signal, all under the control of the galactose-dependent GAL1 promoter. The resulting plasmids were then electroporated into DH10B cells (Gibco/BRL). A total of 7.1 x 10^6 colonies were plated on LB plates containing 100 ug/ml of ampicillin. These *E.coli* were scraped, pooled, and a large scale plasmid prep was performed using the Wizard Maxi Prep kit (Promega, Madison, WI), yielding 3.2mg of supercoiled plasmid DNA.

WI38 Cell cDNA Screening Results:

1×10^6 transformants were obtained on glucose Ura$^-$His$^-$Trp$^-$ plates. These transformants were pooled and resuspended in a solution of 65% glycerol, 10mM Tris-HCl (pH 7.5), 10 mM $MgCl_2$ and stored at -80°C in 1mL aliquots. For screening purposes, aliquots of these were diluted 10-fold into Ura$^-$His$^-$Trp$^-$ CM dropout gal/raff medium (containing 2% galactose, 1% raffinose), which induces the expresssion of the library encoded proteins, and incubated at 30°C for 4 hours. 12×10^6 colony forming units (CFUs) were then plated on standard 10cm galactose X-Gal Ura$^-$His$^-$Trp$^-$Leu$^-$ plates at a density of 2×10^5 CFU/plate. After three days at 30°C, about 1,000 colonies were formed (Leu$^+$) and of those, sixty-four colonies were LacZ$^+$. In order to test if the Leu$^+$/LacZ$^+$ phenotype was due to the library-encoded protein, the galactose dependency of the phenotype was tested. Expression of the library-encoded proteins was turned off by growth on glucose Ura$^-$His$^-$Trp$^-$ master plates and then retested for galactose-dependency on glucose Ura$^-$His$^-$Trp$^-$Leu$^-$, galactose Ura$^-$His$^-$Trp$^-$Leu$^-$, glucose X-Gal Ura$^-$His$^-$Trp$^-$, and galactose X-Gal Ura$^-$His$^-$Trp$^-$ plates. Of these, 32 colonies showed galactose-dependent growth on Leu$^-$ plates and galactose-dependent blue color on X-Gal-containing medium (LacZ$^+$ phenotype). Total yeast DNA was prepared from these colonies according to the method described previously (Hoffman and Winston, 1987). In order to analyze the cDNA sequences, PCR reactions were performed using the above yeast DNA as a template and oligo primers specific for the vector pJG4-5, flanking the cDNA insertion point. PCR products were purified (Qiagen PCR purification kit), subjected to restriction digest with the enzyme HaeIII, run on 1.8% agarose gels, and the restriction patterns compared. Similar and identical restriction patterns were grouped and representatives of each group were sequenced and compared to Genbank and other databases to identify any sequence homologies.

One clone of unique sequence ("2DD") and three clones with identical sequence ("3DD") were isolated and showed no signficant sequence homologies compared to Genbank and other databases. Additionally, four other clones ("20DD") with identical sequence to a portion of human insulin-like growth factor binding protein-5 (Shunichi Shimasaki *et al.*, J. Biol. Chem. *266*:10646-10653 (1991)) were isolated. The clones "2DD," "3DD" and "20DD" were chosen for further analysis. Library vector pJG4-5 containing these clones sequences were rescued from yeast by

transforming the total yeast DNAs into the *E. coli* strain KC8 and selecting for growth on Trp-ampicillin plates. These putative TNFR1 interacting proteins were then tested further for specificity of interaction with the TNF-R1-DD by the reintroduction of JG4-5 clone into EGY48 derivatives containing a panel of different baits, including bicoid, the cytoplasmic domain of the IL-1 receptor, and TNF-R1-DD. The above clones were found to interact only with the TNF-R1-DD. The interaction between these clones and TNF-R1-DD was thus judged to be specific.

U937 cDNA Screening Results:

A U937 cDNA library was also constructed and screened as described above. 1,020 Leu+ colonies were found and of those, 326 colonies were also LacZ+. 62 colonies of these Leu+/LacZ+ colonies showed a galactose-dependent phenotype.. One of these clones, 1TU, encodes a novel sequence. Interestingly, two clones, 15TU and 27TU, encode related or identical sequences, except that 27TU contains about 864 additional nucleotides (or about 288 amino acids) at the 5' end. 15/27TU also encode a novel sequence.

EXAMPLE 2
EXPRESSION OF THE TNF-R1-DD ligand PROTEIN

cDNAs encoding TNF-R intracellular ligand proteins were released from the pJG4-5 vector with the appropriate restriction enzymes. For example, EcoRI and XhoI or NotI and XhoI were used to release cDNA from clone 2DD and clone 20DD. Where the restriction sites were also present in the internal sequence of the cDNA, PCR was performed to obtain the cDNA. For example, the cDNA fragment encoding "clone 3DD" was obtained through PCR due to the presence of an internal XhoI site. These cDNAs were then cloned into various expression vectors. These included pGEX (Pharmacia) or pMAL (New England Biolabs) for expression as a GST (Glutathione-S-transferase) or MBP (maltose binding protein) fusion protein in E. coli, a pED-based vector for mammalian expression, and pVL or pBlueBacHis (Invitrogen) for baculovirus/insect expression. For the immunodetection of TNF-R intracellular ligand expression in mammalian cells, an epitope sequence, "Flag," was inserted into the translational start site of the pED vector, generating the pED-Flag vector. cDNAs

were then inserted into the pED-Flag vector. Thus, the expression of cDNA from pED-Flag yields a protein with an amino terminal Met, followed by the "Flag" sequence, Asp-Tyr-Lys-Asp-Asp-Asp-Asp-Lys. Standard DEAE-Dextran or lipofectamine methods were used to transfect COS or CHO dukx cells. Immunodetection of Flag-tagged proteins was achieved using the M2 antibody (Kodak). Moreover, an immunoaffinity column using the M2 antibody, followed by elution with the "Flag" peptide, can be used for the rapid purification of the flag-tagged protein. Similarly, affinity purification of GST- , MBP- or His-tagged fusion proteins can be performed using glutathione, amylose, or nickel columns. Detailed purification protocols are provided by the manufacturers. For many fusion proteins, the TNF-R intracellular ligand can be released by the action of thrombin, factor Xa, or enterokinase cleavage. In the case where highly purified material is required, standard purification procedures, such as ion-exchange, hydrophobic, and gel filtration chromatography will be applied in addition to the affinity purification step.

Figs. 1 and 2 depict autoradiographs demonstrating the expression of TNF-R1-DD ligand proteins in yeast and mammalian cells. Fig. 1 shows the results of expression of isloated clones of the present invention in yeast. EGY48 was transformed with pJG4-5 containing clone 2DD, 3DD or 20DD. Cells were then grown overnight in the galactose/raffinose medium. Cell lysates were prepared and subject to 4-20% SDS gel electrophoresis, followed by Western blot analysis using anti-HA antibody (12CA5, Boehringer Mannheim, Indianapolis, IN). Fig. 2 shows the results of expression of Flag-2DD and Flag-20DD in COS cells. COS cells were transfected with either pED-Flag (Vector control), Flag-2DD or Flag-20DD plasmid by the lipofectamine method. Thirty µg of each cell lysate were prepared and subjected to 4-20% SDS gel electrophoresis, followed by Western blot analysis using M2 antibody (Kodak). The bands in the Flag-2DD and Flag-20DD lanes indicate significant expression of the respective TNF-R1-DD ligand proteins.

EXAMPLE 3
ASSAYS OF TNF-R DEATH DOMAIN BINDING

Two different methods were used to assay for TNF-R1-DD ligand protein activity. The first assay measures binding in the yeast strain in "interaction trap," the system used here to screen for TNF-R1-DD interacting proteins. In this system, the expression of reporter genes from both LexAop-Leu2 and LexAop-LacZ relies on the interaction between the bait protein, in this case TNF-R1DD, and the prey, the TNF-R intracellular ligand. Thus, one can measure the strength of the interaction by the level of Leu2 or LacZ expression. The most simple method is to measure the activity of the LacZ encoded protein, β-galactosidase. This activity can be judged by the degree of blueness on the X-Gal containing medium or filter. For the quantitative measurement of β-galactosidase activity, standard assays can be found in "Methods in Yeast Genetics" Cold Spring Harbor, New York, 1990 (by Rose, M.D., Winston, F., and Hieter, P.).

The second assay for measuring binding is a cell-free system. An example of a typical assay is described below. Purified GST-TNF-R1-DD fusion protein (2 ug) was mixed with amylose resins bound with a GST-TNF-R1-DD intracellular ligand for 2 hour at 4°C. The mixture was then centrifuged to separate bound (remained with the beads) and unbound (remained in the supernatant) GST-TNF-R1-DD. After extensive washing, the bound GST-TNF-R1-DD was eluted with maltose and detected by Western blot analysis using a GST antibody. The TNF-R1-DD or the intracellular ligand can also be immobilized on other solid supports, such as on plates or fluorobeads. The binding can then be measured using ELISA or SPA (scintillation proximity assay).

EXAMPLE 4
CHARACTERIZATION OF TNF-R DEATH DOMAIN LIGAND PROTEIN

Mapping the interaction site in TNF-R1

Many of the key amino acids for TNF-R signaling have been determined by site-directed mutagenesis (Tataglia *et al.*, Cell *74*:845-853 (1993). These amino acids are conserved between TNF-R and the Fas antigen, which is required for

mediating cytotoxicity and other cellular responses. In order to test if the TNF-R intracellular proteins interact with these residues, the following mutations were constructed: F345A (substitution of phe at amino acid 345 to Ala), R347A, L351A, F345A/R347A/L351A, E369A, W378A and I408A. The ability of the mutant protein to interact with the intracellular ligand in the "interaction trap" system was tested.

Effect on the TNF-mediated response

The effect of the TNF-R intracellular ligands on the TNF-mediated response can be evaluated in cells overexpressing the ligands. A number of TNF-mediated responses, including transient or prolonged responses, can be measured. For example, TNF-induced kinase activity toward either MBP (myelin basic protein) or the N-terminus (amino acids 1-79) of c-jun can be measured in COS cells or CHO cells either transiently or stably overexpressing clone 2DD, 3DD or clone 20DD. The significance of these ligand proteins in TNF-mediated cytotoxicity and other cellular responses can be measured in L929 or U937 overexpressing cells. Alternatively, other functional assays, such as the induction of gene expression or PGE_2 production after prolonged incubation with TNF, can also be used to measure the TNF mediated response. Conversely, the significance of the TNF-R1-DD ligand proteins in TNF signaling can be established by lowering or eliminating the expression of the ligands. These experiments can be performed using antisense expression or transgenic mice.

Enzymatic or functional assays

The signal transduction events initiated by TNF binding to its receptor are still largely unknown. However, one major result of TNF binding is the stimulation of cellular serine/threonine kinase activity. In addition, TNF has been shown to stimulate the activity of PC-PLC, PLA_2, and sphingomyelinase. Therefore, some of the TNF-R1-DD ligand proteins may possess intrinsic enzymatic activity that is responsible for these activities. Therefore, enzymatic assays can be performed to test this possibility, particularly with those clones that encode proteins with sequence homology to known enzymes. In addition to enzymatic activity, based on the sequence homology to proteins with known function, other functional assays can also be measured.

EXAMPLE 5
ISOLATION OF FULL LENGTH CLONES

In many cases, cDNAs obtained from the interaction trap method each encode only a portion of the full length protein. For example, based on identity and sequence and the lack of the initiating methionine codon, clones 2DD, 3DD and 20DD apparently do not encode full length proteins. Therefore, it is desirable to isolate full length clones. The cDNAs obtained from the screening, such as clone 2DD, are used as probes, and the cDNA libraries described herein, or alternatively phage cDNA libraries, are screened to obtain full length clones in accordance with known methods (see for example, "Molecular Cloning, A Laboratory Manual", by Sambrook et al., 1989 Cold Spring Harbor).

EXAMPLE 6
ANTIBODIES SPECIFIC FOR TNF-R INTRACELLULAR LIGAND PROTEIN

Antibodies specific for TNF-R intracellular ligand proteins can be produced using purified recombinant protein, as described in Example 2, as antigen. Both polyclonal and monoclonal antibodies will be produced using standard techniques, such as those described in "Antibodies, a Laboratory Manual" by Ed Harlow and David Lane (1988), Cold Spring Harbor Laboratory.

EXAMPLE 7
CHARACTERIZATION OF CLONES 1TU AND 15/27TU

Specificity of Interaction

The specificity of clones 1TU, 15TU and 27TU was tested using a panel of baits. The ability of these clones to bind the TNF-R death domain was compared to their binding to the intracellular domain of the second TNF-R (TNF-R $p75_{IC}$), the entire intracellular domain of TNF-R (TNF-R $p55_{IC}$), the death domain of the fas antigen (which shares 28% identity with TNF-R-DD) (Fas_{DD}), the *Drosophila* transcription factor bicoid, and a region of the IL-1 receptor known to be critical for signalling ($IL\text{-}1R_{477\text{-}527}$). As shown in Table 1, none of these clones interacted with TNF-R $p75_{IC}$ or

Fas_{DD}, and only 1TU interacted with bicoid. In contrast, both 1TU and 15TU bound the cytoplasmic domain of the p55 TNF-R, as well as residues 477-527 of the IL-1R. 27TU interacted relatively weakly with these sequences.

Table 1

clone	TNF-R_{DD}	TNF-R $p75_{IC}$	TNF-R $p55_{IC}$	Fas_{DD}	bicoid	IL-1R (477-527)
1TU	+++	-	+++	-	++	+++
15TU	+++	±	+++	-	-	++
27TU	+++	-	+	-	-	+

Interaction with Amino Acids Critical for Signalling

The ability of each clone to interact with four single-site mutations in the TNF-R death domain (each known to abolish signalling) was measured. As shown in Table 2, each of the clones interacted less strongly with the death domain mutants than with the wild type death domain, suggesting that these clones may bind critical residues *in vivo*.

Table 2

clone	TNF-R_{DD}	F345A	L351A	W378A	I408A
1TU	+++	+	++	++	+
15TU	+++	+	+	++	++
27TU	+++	+	+	±	++

Expression of 1TU, 15TU and 27TU

Fig. 3 depicts an autoradiograph demonstrating the expression of clones 1TU, 15TU and 27TU in yeast (A) and COS cells (B).

In (A): EGY48 was transformed with pJG4-5 containing clones 1TU, 15TU or 27TU. Cells were then grown overnight in galactose/raffinose medium. Cell lysates were prepared and subjected to 4-20% SDS gel electrophoresis, followed by Western blot analysis using anti-HA antibody (12CA5, Boehringer Mannheim).

In (B): COS cells were transfected with pED-Flag containing clones 1TU, 15TU and 27TU. Cell lysates were prepared and analyzed by Western blot using anti-Flag antibody (M2, Kodak).

Specific Binding of 1TU and 27TU to TNF-R1-DD

The interaction of 1TU and 27TU with TNF-R1-DD was tested using purified bacterially expressed fusion proteins. As shown in Fig. 4, MBP fusion proteins containing 1TU or 27TU bound only to TNF-R1-DD expressed as a GST fusion protein, but not to GST protein alone. In the control experiment, MBP protein did not bind either GST or GST/TNF-R1-DD. These results indicate that 1TU and 27TU bound specifically to the TNF-R1 death domain *in vitro*, confirming the data obtained in the interaction trap.

15TU and 27TU Activation of JNK Activity

The jun N-terminal kinase (JNK) is normally activated within 15 min of TNF treatment in COS cells. 15TU and 27TU were cotransfected with an epitope tagged version of JNK, HA-JNK, in duplicate. After TNF treatment, JNK was immunoprecipitated with anti-HA antibody and JNK activity was measured in immunoprecipitation kinase assays, using GST-c-jun (amino acids 1-79) as substrate). Reactions were electrophoresed on SDS-PAGE. As shown in Fig. 5, transfection of 15TU and 27TU, but not vector alone, into COS cells activated JNK even in the absence of TNF, suggesting that these clones are involved in signal transduction of TNF and the pathway leading to JNK activation *in vivo*.

EXAMPLE 8
ISOLATION, EXPRESSION AND ASSAY OF CLONE 3TW

Clone 3TW was isolated from the WI38 cDNA library using clone 3DD as a porbe. Clone 3TW was expressed. Fig. 6 is an autoradiograph which demonstrates expression of 3TW (indicated by arrow).

An antisense oligonucleotide was derived from the sequence of clone 3TW. The antisense oligonucleotide was assayed to determine its ability to inhibit TNF-induced $cPLA_2$ phosphorylation. Fig. 7 depicts the results of that experiment. Activity of the anitsense oligonucleotide (3TWAS) was compared with the full-length clone (3TWFL), Flag-3TW full length (3TWFLflag) and pED-flag vector (pEDflag). The antisense oligonucleotide inhibited phosphorylation.